Experimental Section

General protocol: The 2-deoxythioglycoside (0.1 mmol), alcohol (1.5 equiv), and DTBMP (4.5 equiv) were dissolved in CH_2Cl_2 (1.0 mL) under argon, using light-protected glassware. After the mixture had been stirred for 1.5 h at 23 °C over molecular sieves (4 Å, <5 microns, freshly activated), powdered AgPF₆ (3–4 equiv) was added at O °C. When the reaction was complete (0.5–2 h), pyridine (50 equiv) was added, and the mixture was stirred for a further 0.5 h. Filtration (celite pad, diethyl ether/n-hexane (1:4)), concentration, and chromatographic purification provided the 2-deoxyglycoside.

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in italics: IR (film): $\tilde{v}_{\rm max}\!=\!2929,\ 1757,\ 1734,\ 1654,\ 1618,\ 1560,\ 1515,$ 1448, 1396, 1247, 1124, 739, 618 cm⁻¹; ¹H NMR (600 MHz, CDCl₃): δ = 9.04 (d, J = 7.8 Hz, 1 H; NH), 8.93 (s, 1 H; H-1"), 7.23 and 6.84 (d \times $2, J = 8.6 \text{ Hz}, 1 \text{ H} \times 2; \text{ PMBM}), 7.21 \text{ (d, } J = 8.1 \text{ Hz}, 1 \text{ H}; \text{ H} - 4'), 7.19 \text{ (d, }$ J = 8.0 Hz, 1 H; H-5'), 7.04 (s, 1 H; H-4''), 6.80 (s, 1 H; H-5''), 6.34 (m,1 H; Allyl), 6.07 (d, J = 2.2 Hz, 1 H; H-12), 5.84 (m, 1 H; Alloc), 5.73 (m, 1H; H-7'), 5.61 (br dd, J = 2.2, 3.5 Hz, 1H; H-11), 5.49 and 5.29 (br dd \times 2, J = 1.3, 17.1 Hz, 1 H \times 2; Allyl), 5.41 and 5.23 (br dd \times 2, J = 1.0, 10.3 Hz, 1 H × 2; Alloc), 5.40 (br s, 1 H; H-1'''), 4.81 and 4.78 $(d \times 2, J = 7.0 \text{ Hz}, 1 \text{ H} \times 2; \text{PMBM}), 4.71 \text{ (sept, } 1 \text{ H}; \text{H}-10''), 4.53 - 4.48$ (brs + m, 6H; PMBM + Allyl + Alloc), 4.26 (dd, J = 12.6, 6.0 Hz,1H; $H_{a/b}$ -14), 4.22 (d, J = 8.6 Hz, 1H; $H_{a/b}$ -8), 4.19 (d, J = 3.6 Hz, 1H; H-10), 4.17 (d, J = 12.6 Hz, 1 H; $H_{b/a}$ -14), 4.08 (d, J = 8.6 Hz, 1 H; $H_{b/a}$ -8), 4.06 (s + m, 4H; CH₃-14" + H-5""), 4.03 (d, J = 6.2 Hz, 1H; H-13), 3.91 (s, 3H; CH_3 -13"), 3.72 (s, 3H; PMBM), 3.26 (d, J = 9.3 Hz, 1H; H-4''''), 3.13 (dd, J = 14.7, 4.7 Hz, 1 H; $H_{a/b}$ -8'), 3.09 and 3.00 (br d and $d, J = 16.4 Hz, 1 H \times 2; CH_2-5), 2.62 (dd, J = 14.7, 11.6 Hz, 1 H; H_{b/a}-8'),$ $2.14 (d, J = 14.4 Hz, 1H; H_{a/b}-2''''), 1.85 (dd, J = 14.4, 4.3 Hz, 1H; H_{b/a}-1.85 Hz,$ 2""), 1.49 and 1.43 (s \times 2, 3 H \times 2; acetonide), 1.46 and 1.45 (d \times 2, J = 5.9 Hz, $3 \text{ H} \times 2$; CH_3 -12'' and CH_3 -13''), $1.33 \text{ (d, } J = 6.3 \text{ Hz, } 3 \text{ H; } \text{CH}_3$ -6""), 1.21 (s, 3H; CH₃-7""), 1.00 (m, 18H; TES × 2), 0.68 (m, 12H, TES \times 2); HR-MS (MALDI-TOF) m/z for $C_{76}ClH_{101}N_2NaO_{20}Si_2^{+}$ $[M+Na]^+$: calcd: 1475.6052, found: 1475.6000.

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H₂O₂-Dependent Fe-Catalyzed Oxidations: Control of the Active Species**

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In memory of Olivier Kahn

The reaction of ferrous ion with hydroperoxides has been investigated for more than a century (Fenton chemistry), and yet the mechanism has still not been satisfactorily rationalized. The nature of the reactive species has always been a matter of debate, oscillating between hydroxyl or alkoxyl

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radicals and an oxene species.[1,2] Moreover, the discovery that non-heme-iron proteins can oxidize inert substrates (e.g., alkanes) selectively has stimulated chemists to reinvestigate the reaction of non-heme-iron complexes with dioxygen- and oxygen-derived oxidants.[3] In fact, dinuclear, methane monooxygenase, [4] and mononuclear non-hemeiron enzymes such as isopenicillin N synthase, 2-oxo-glutarate-dependent enzymes, and pterin-dependent phenylalanine, tyrosine, tryptophan hydroxylases are proposed to carry out a two-electron oxidation via an Fe^{IV}-oxo moiety.^[5] Herein, we report on the oxidation mechanism by using iron complexes of the ligand N,N'-bis(pyridin-2-ylmethyl)-N,N'bis(3,4,5-trimethoxybenzyl)ethane-1,2-diamine (L) as a substrate. [6] This ligand displays several potential oxidation sites, in particular the methylene groups and phenyl rings from benzylic moieties; the latter are known to be very reactive towards hydroxyl radicals, leading to the formation of phenol.^[7] Herein we compare the catalytic properties of two complexes, $[Fe^{II}LCl_2] \cdot 0.25H_2O$ (1 · 0.25 H₂O) and $[Fe^{II}L(CH_3CN)_2(CIO_4)_2] \cdot 3H_2O$ (2(CIO₄)₂ · 3H₂O), during oxidation of ligand L (Scheme 1) and of various substrates by H₂O₂ in acetonitrile. With such probes we demonstrate that complex 1 behaves as a typical Fenton reagent, whereas complex 2 functions through metal-based mechanisms. This

OMe Me X = CI 1 $X = CH_3CN$ 2 equiv H₂O₂ 2 2 equiv H₂O₂ .OMe MeC MeO Extraction Extraction OMe OMe L1 OMe LOH

Scheme 1. Ligand oxidation of complexes ${\bf 1}$ and ${\bf 2}$ to give LOH and L1, respectively.

shows that for this class of iron complexes the reactivity can be tuned by modifying the simple monodentate ligand.

The synthesis of complexes 1 and 2 started with mixing equimolar amounts of FeCl₂·4H₂O or Fe(ClO₄)₂·6H₂O, respectively, and the ligand L in CH₃OH under argon. Addition of diethyl ether to the CH₃OH solution afforded in both cases a yellow powder. Slow diffusion of diethyl ether into a solution of these powders in acetonitrile afforded crystals suitable for X-ray structure analysis (Figure 1).[8] The crystal structures of 1 · CH₃CN and 2(PF₆)₂ · 2.5 CH₃CN · 0.25 CH₃OH · 0.5 H₂O reveal that in each case the Fe center is octahedrally coordinated; the coordination differs with respect to the nature of the two ligands that occupy the two cis-positioned open sites of the Fe^{II} coordination sphere: in compound 1 they are filled by chloride ions, whereas in 2 they are filled by acetonitrile solvent molecules. This difference confers a different spin state to the ferrous ion in the solid state only (shown by the longer Fe-N distances in 1 than in 2). Complexes 1 and 2 displayed significantly different

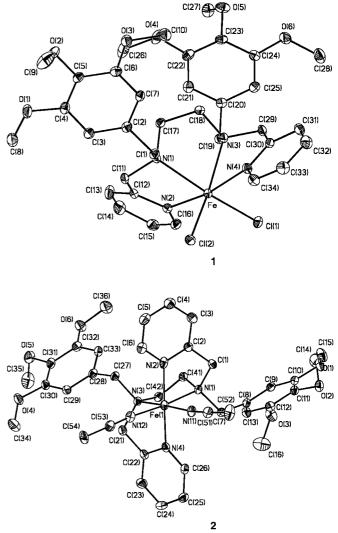


Figure 1. ORTEP representation of complexes **1** and **2**. Hydrogen atoms have been omitted for clarity. Selected bond lengths [Å]: for **1**: Fe-N(1) 2.3272(7), Fe-N(2) 2.2120(6), Fe-N(3) 2.3012(6), Fe-N(4) 2.2228(6); for **2**: Fe(1)-N(11) 1.939(4), Fe(1)-N(12) 1.942(4), Fe(1)-N(2) 1.986(4), Fe(1)-N(4) 1.988(4), Fe(1)-N(1) 2.055(4), Fe(1)-N(3) 2.061(4).

resonance signals in their 1H NMR spectra in acetonitrile between $\delta = 160$ and -20 (data not shown).

When an excess of H₂O₂ was added to a solution of complex 1 (1 mm) in acetonitrile under argon, the solution immediately turned blue. Titration studies showed that only two equivalents of hydrogen peroxide were required to achieve maximal intensity of a new transition at 740 nm. The new species was characterized as a mononuclear phenolatoiron(III) complex [Fe^{III}(LO)Cl₂] (Scheme 1) based on the following spectroscopic signatures: 1) the transition at 740 nm was assigned to a charge-transfer band from an oxygen atom of the phenolato ligand to an iron(III) center; [9] 2) the positiveion mode ESI mass spectrum of the resulting solution displayed only one molecular peak at m/z 708 (100%) attributed to $[Fe(L-H)Cl+O]^+$ whose isotopic pattern was in agreement with the theoretical one; 3) an isotropic signal was observed at g = 4.3 in the EPR spectrum of the resulting solution.

The oxidized ligand was then extracted and characterized ^[10] by ESI-MS and ¹H NMR analysis as N-(2-hydroxy-3,4,5-trimethoxybenzyl)-N,N'-bis(pyridin-2-ylmethyl)-N'-(3,4,5-trimethoxybenzyl)ethane-1,2-diamine which results from the hydroxylation at the *ortho* position of one phenyl ring of the ligand. ^[11] HPLC titration showed that more than $70(\pm 10)$ % of the ligand L had been transformed into LOH and only $6(\pm 5)$ % of the starting ligand was found unconverted. ^[12] Mass spectrometric analysis of the modified ligand LOH obtained when $H_2^{18}O_2$ was used as an oxidant established that the oxygen atom incorporated came exclusively from the oxidant. Control experiments in the presence of $H_2^{16}O$ in the $H_2^{18}O_2$ -dependent reaction did not alter the yield of ¹⁸O-atom incorporation. Aromatic hydroxylation of the ligand in iron complexes has been previously reported. ^[6, 13]

In contrast, addition of H_2O_2 to complex $2\cdot (ClO_4)_2$ under the same conditions as for 1 did not result in any intense color change. However, extraction of the organic material from the reaction mixture and analysis by HPLC led to the recovery of $65(\pm 10)$ % of the unconverted ligand L and the isolation of a new modified ligand L1 in $14(\pm 5)$ % yield (see Scheme 1). L1 was characterized by ESI-MS and HNMR analysis solution as N_iN^i -bis(pyridin-2-ylmethyl)-N-(3,4,5-trimethoxybenzyl)ethane-1,2-diamine which resulted from an N-dealkylation of the L ligand. No evidence for the formation of LOH in this reaction was obtained. Clearly, the nature of the two monodentate ligands (Cl vs. CH_3CN) influenced the selectivity of the ligand oxidation, suggesting the participation of different oxidative species.

To distinguish between these reactive species, the catalytic properties of the $1/H_2O_2$ or $2/H_2O_2$ combination towards external substrates was studied.^[17] Only 2 led to catalytic and stereoselective alkane hydroxylation under anaerobic conditions. For example, the reaction of *cis*-1,2-dimethylcyclohexane with $2/H_2O_2$ led to the catalytic formation of *cis*-1,2-dimethylcyclohexanol and *trans*-1,2-dimethylcyclohexanol in a 10:1 ratio (yield based on the oxidant 20%), whereas the analogous reaction with the $1/H_2O_2$ combination led to an equimolar mixture of *cis*- and *trans*-1,2-dimethylcyclohexanol (yield 7%). Furthermore, catalytic oxidation of cyclohexane to give cyclohexanol and only minor amounts of cyclohex

anone (yield based on the oxidant 38%; alcohol/ketone 8/1) occurred only with complex 2, whereas equimolar amounts of alcohol and ketone were detected with complex 1 as the catalyst (yield based on the oxidant 6%). The origin of the oxygen atom incorporated into cyclohexane has been determined in both cases.[18] When 10 equivalents of H₂18O₂ were mixed with 2 in the presence of 1000 equivalents of H₂¹⁶O, $47(\pm 3)\%$ of the cyclohexanol was ¹⁸O-labeled. This was confirmed when ¹⁸O-labeled water and unlabeled H₂O₂ were used. In contrast, with complex 1 and H₂¹⁸O₂, cyclohexanol was fully labeled with the oxygen atom from the oxidant; no exchange with water was observed. Finally, only complex 2 was capable of epoxidizing cyclooctene (30% yield based on the oxidant) and cyclohexene (20% yield based on the oxidant) substrates.^[19] Notably, in the case of cyclohexene, allylic oxidation products were the major products regardless of the catalyst (only 2-cyclohexen-1-ol was formed; yield 10%).

All these data taken together imply two different oxidative pathways depending upon the catalyst structure. In the case of complex 1, the quasi-quantitative hydroxylation of the pendant phenyl ring of the ligand, the origin of the oxygen atom from the oxidant, the poor (stereo)selectivity during alkane oxidation, and the lack of epoxidation strongly suggest that freely diffusing OH is the active species. [20, 21] In contrast, in the case of complex 2, the high stereoselectivity, the solvent oxygen atom exchange, the oxidation of the closest C-H bond of the ligand (the methylene group of the benzyl ring), and the reasonable yield of epoxides can be interpreted with the involvement of an iron-oxo complex as an active species, allowing a better control of the substrate radicals ("caged radicals") by the metal center. The participation of free hydroxyl radicals when catalyst 2 was used is thus unlikely. The presence of products labeled with oxygen atoms from both water and the oxidant has already been observed in oxidations dependent on heme- and non-heme-iron complexes and has been explained by a postulated "oxo-hydroxo tautomerism" of the Fe^{IV} or Fe^V=O intermediate. [22-25]

One possible explanation for the difference between 1 and 2 is the presence of labile sites only in 2, which thus allow the binding of H_2O_2 to the metal center. Complex 1 would thus react with H_2O_2 through an outer-sphere electron transfer resulting in formation of OH'. In contrast, with complex 2 an iron – peroxo species, similar to peroxo complexes previously observed during H_2O_2 -dependent reactions, can be transiently formed as a precursor for a metal-based oxidative species. [26] The nature of this species remains to be established. We favor a homolytic cleavage of the FeO–OH bond, leading to a caged [Fe^{IV}=O OH'] radical pair.

This work further supports the notion that high-valent iron-oxo species (and not only OH radicals) can be generated and used as active species in non-heme-iron catalysis. Furthermore, it shows that the balance between different catalytic pathways for the oxidation (OH vs. metalbased oxidant) can be tuned through the manipulation of the coordination sphere of the ferrous ion.

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- [10] The resulting solution was concentrated and diluted in water in the presence of excess dithionite and bathophenanthroline. The organic material was extracted from the solution by using dichloromethane and choloroform.
- [11] Comparisons between ¹H and ¹³C NMR spectra of L and LOH allowed us to unambiguously attribute the inserted oxygen position. In fact, the signal at $\delta = 6.63$ corresponding to the o-H for L is replaced by two signals at $\delta = 6.63$ and 6.42 in a 2:1 ratio for LOH. A similar trend is observed in the ¹³C NMR spectrum (for the o-C of the phenyl ring signals appear at $\delta = 108.5$ and 105.5 for LOH, but only at $\delta = 105.0$ for L). ESI-MS for LOH (CH₂Cl₂; positive-ion mode): m/z: 619 (100 %) for [LOH + H⁺].
- [12] The HPLC titration has been carried out on authentic pure samples of L, L1, and LOH obtained by extraction of the reaction mixture at

- higher concentration. Other minor products have been detected but their structures remain unknown.
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- [14] The reaction mixture contains both mononuclear Fe^{II} and Fe^{III} L and L1 complexes.
- [15] ¹H NMR (300 MHz, CD₃CN, 25 °C): δ = 2.66 (m, 4H; NCH₂CH₂N), 3.52 (s, 2H; CH₂Bz), 3.62 (s, 3H; OCH₃), 3.67 (s, 2H; CH₂py), 3.69 (s, 6H; OCH₃), 3.72 (s, 2H; CH₂py), 6.63 (s, 2H; o-H_{ph}); 7.13 (td, 2H; m-H_{py}); 7.23 (d, 1H; p-H_{py}), 7.43 (d, 1H; p-H_{py}), 7.61 (t, 2H; m-H_{py}), 8.41 (dd, 2H, o-H_{py}). ESI/MS (CH₂Cl₂; positive-ion mode): m/z: 423 (100 %) for [L1 + H⁺].
- [16] Accordingly, the pendant product of the N-dealkylation of the ligand, 3,4,5-trimethoxybenzaldehyde, has been quantitatively detected.
- [17] Experimental conditions: complex/alkane/H₂O₂ 1/1100/10 in acetonitrile inside a glove box (dioxygen content less than 1 ppm) using syringe pump conditions; delivering time = 30 min [complex] = 0.7 mm.
- [18] The isotope ratio in cyclohexanol has been analyzed by GC/MS on a reaction mixture directly after reaction. Cyclohexanol was not labeled in the presence of ¹⁸O water under standard conditions.
- [19] No 1,2-cyclohexanediol was detected.
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A Route to a Germanium – Carbon Triple Bond: First Chemical Evidence for a Germyne**

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Since the isolation of the first disilene, a compound containing a silicon-silicon double bond, in 1981,^[1] many stable compounds with a double bond involving the Group 14 elements Si, Ge, and Sn have been synthesized.^[2] In contrast, compounds containing a triple bond to a Group 14 element other than C remained unknown. Power^[3] described recently a lead analogue of an alkyne (ArPbPbAr) but more consistent with a diplumbylene form. However, some compounds with triple bonds to Si were identified by spectroscopy or by

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